AN SAR STUDY FOR THE NON-PEPTIDE SUBSTANCE P RECEPTOR (NK₁) ANTAGONIST, CP-96,345.¹

William Howson,* Julie Hodgson, Reg Richardson, Lesley Walton, Steve Guard and Keith Watling.

Parke-Davis Neuroscience Research Centre, Addenbrookes Hospital Site, Hills Road, Cambridge CB2 2QB, UK.

(Received 4 February 1992)

Abstract : Results from an SAR study around the novel, non-peptide substance P receptor NK_1 antagonist, (±)CP-96,345 (1) are described. The importance of the 2° nitrogen and the aromatic moleties are clarified.

The mammalian tachykinin neuropeptides, substance P, neurokinin A, neurokinin B, neuropeptide K and neuropeptide Y are widely distributed throughout the central and peripheral nervous systems, and all contain a common C-terminal sequence, -Phe-xxx-Gly-Leu-MetNH2.2 Of these substance P, isolated in 1931 by von Euler and Gaddum, 3 but characterized only more recently in 1970 by Chang and Leeman, has been the most extensively studied. There is good evidence to support the existence of at least three distinct tachykinin receptor types, 5 termed NK1, NK2 and NK3, which bind preferentially substance P, neurokinin A and neurokinin B, respectively. These receptors mediate a variety of biological effects of the tachykinin neuropeptides, including smooth muscle contraction, vasodilation, bronchoconstriction, salivary secretion and neuronal excitation, and thus represent major targets for drug research in this area. detailed understanding of the physiological roles of neuropeptides, both in the periphery and the central nervous system, has not been possible because of the lack of selective tachykinin receptor agonists and antagonists possessing high affinity and good in vivo activity. However, Snider et al. (Pfizer Inc.) have recently described a novel, high affinity, non-peptide antagonist of the NK1 receptor, Lowe et al (Pfizer Inc.) have subsequently reported the

CP-96,345. Lowe et al (Pfizer Inc.) have subsequently reported the preparation and radiolabelling of CP-96,345 along with its affinity and that of two analogues (the 2-Cl and unsubstituted benzyl derivatives, compounds 2 and 6 respectively in this study) for the NK_1 receptor in bovine caudate using [3H]SP as the radioligand. Presented here are the results of a study to determine the importance of various functional groups present in racemic ((4)CP-96,345 (1).

The compounds studied are shown in table 1 and their syntheses outlined in Scheme 1; all compounds are racemates. Compounds 1-8 were prepared by the method of Lowe, briefly this was as follows. A base

catalysed condensation of 3-quinuclidinone with benzaldehyde gave the enone 9. Regioselective addition of phenylmagnesium bromide to 9 with ${\rm CuCl_2}$ catalysis gave the ketone 10. This could be elaborated to the final compounds 1-8 by stereoselective reductive amination with the appropriate amine and 9-BBN. Compound 11 was prepared by a similar reductive amination of ketone 12.8

Hydrogenolysis of 6 gave the 1° amine $13,^{9}$ treatment of this with 2-chlorobenzoic acid activated with DCC gave the amide 14.

The ether 15 was obtained by stereoselective reduction of the ketone $10,^9$ followed by regiospecific alkylation of the alcohol 16 with 2-chlorobenzylbromide using powdered KOH in DMSO.

The trans isomer of 1, compound 17 was prepared using the methodology of Warawa and Mueller. 9

Scheme 1

$$CH_2O$$
 CH_2O
 CH_2O

a PhMgBr, CuCl₂, THF, b NH(CH₂)_n, H*, Tol,
$$\Delta$$
, then 9- BBN, THF, c When n = 1, X = H: H₂ / Pd/C, HCl, MeOH, Cl d CO_2H , DCC, CH₂Cl₂, e H₂ / Pd / C, HCl, EtOH; 1 A I (OPr¹) 3, Pr¹ OH, Δ ; g CH_2Br , KOH, DMSO OCH₃

Table 1: Binding of(+/-) CP-96,345(1)analogues and standards to NK_1 sites in guinea pig and rat cerebral cortex membranes

$$\bigcap_{N}^{R}$$

No.	R ₁	R ₂	NK ₁ Binding IC ₅₀ (nM) Guinea Pig Rat cerebral cortex cerebral cortex		IC ₅₀ ratio Rat / Guinea Pig	
1	NHCH ₂ (2-CH ₃ OPh)	CH(Ph) ₂	1.2(-0.2,+0.3) ^a	120(-15,+17)	100	
2	NHCH ₂ (2-CIPh)	CH(Ph) ₂	8.5(-1.7,+2.0)	110(-15,+17)	13	
3	NHCH ₂ (2-CF ₃ Ph)	CH(Ph) ₂	33(-2,+3)	740(-120,+150)	22	
4	NHCH ₂ (2-FPh)	CH(Ph) ₂	37(-12,+17)	990(-240,+310)	27	
5	NHCH ₂ (2-CH ₃ Ph)	CH(Ph) ₂	227(-30,+35)	700(-110,+130)	3	
6	NHCH ₂ Ph	CH(Ph) ₂	52(-10,+13)	2000(-500,+660)	38	
7	NH(CH ₂) ₂ (2-CIPh)	CH(Ph) ₂	160(-25,+29)	7200(-1000,+1200)	45	
8	NH(CH ₂) ₃ (2-CIPh)	CH(Ph) ₂	1100(-270,+360)	>10,000	>9	
1 1	NHCH ₂ (2-CH ₃ OPh)	CH ₂ Ph	1510(-380,+580)	>10,000	>7	
1 3	NH ₂	CH(Ph) ₂	>10,000	>10,000	-	
1 4	NHCO(2-CIPh)	CH(Ph) ₂	>10,000	>10,000	-	
1 5	OCH ₂ (2-CH ₃ OPh)	CH(Ph) ₂	150(-34,+44)	~10,000	67	
1 7	trans isome	er of 1	10(-1.3,+1.5)	800(-180,+230)	80	
	Substance P		0.07(-0.01,+0.01)	0.12(-0.01,+0.01)	1.8	
	Neurokinin A		3.5(-1.3,+2.0)	31(-9,+12)	9	
			13(-3,+5)	69(-15,+19)	5	
	Spantide		820(-190,+250)	650(-180,+250)	0.8	

a(geometric mean - , + s.e.m)

The affinities of the above compounds for NK_1 receptor binding sites present in either rat or guinea-pig cerebral cortex membranes were determined according to the method of Lee <u>et al</u>, ¹⁰ with minor modifications. ¹¹ The results are collated in Table 1, together with those obtained for the endogenous tachykinins and the peptide based antagonist $[D-Arg^1, D-Trp^{7,9}, Leu^{11}]SP$ (spantide).

In membranes prepared from guinea-pig cerebral cortex comparison of the affinities of compounds 1-6 highlight the importance of the ortho aromatic substituent, the $2-\text{CH}_3\text{O}$ analogue 1 binding 50-fold better than the unsubstituted compound 6. Attempts to correlate a range of physicochemical properties 12 of the ortho substituents with the affinities of the compounds were unsuccessful (see Table 2).

Table 2: NK₁ Binding data and physicochemical parameters for ortho substituents X

No.	x	Guinea Pig NK ₁ binding (IC ₅₀ , nM)		MR	Es	F	Я
1	сң₀о	1.2	-0.02	7.87	-0.55	0.26	-0.51
2	CI	8.5	0.71	6.03	-0.97	0.41	-0.15
3	CF ₃	33	0.88	5.02	-2.40	0.38	0.19
4	F	37	0.14	0.92	-0.46	0.43	-0.34
6	Н	52	0	1.03	0	0	0
5	CHa	227	0.56	5.65	-1.24	-0.04	-0.13

The benzyl group itself was shown to be highly significant for binding, its removal leading to the inactive 1° amine 13.

The distance between the secondary nitrogen and ortho substituted phenyl was also found to have a considerable influence on binding. As this distance increased in compounds 2, 7, and 8 there was a gradual decrease in affinity.

The significance of the basic secondary nitrogen of the benzylamine moiety was investigated by replacing it with a neutral oxygen atom (15) or reducing its basic character by replacing the adjacent CH₂ with CO (14).

Both of these analogues were considerably less active, the ether 15 some 100-fold less and the amide 14 inactive.

Removal of one of the aromatic rings of the benzylhydryl moiety led to the very weakly active derivative 11 suggesting that either both rings of this group bind to the NK_1 receptor or that its effect on conformation is critical.

Finally, our results indicate that the stereochemistry is important, the trans analogue of 1, compound 17 possessing only one tenth of the affinity.

When examined for their ability to displace the binding of [125 I]-BHSP to NK₁ sites present in rat cerebral cortex these compounds show a similar rank order of affinity, but all are less active. The decrease in affinity varies across the series, the largest difference being observed for the ortho methoxy analogue 1 (100-fold) and the smallest for the ortho methyl analogue 5 (3-fold). These data are in contrast to that observed for neurokinin A, neurokinin B, substance P and spantide which show similar binding affinities for both guinea-pig and rat NK₁ receptors (see Table 1).

Compounds 1-8, 11, 13-15 and 17 were also examined for their ability to displace the binding of [125 I-iodohistidyl]-NKA to NK₂ sites present in hamster urinary bladder membranes 13 and [3 H]-Senktide to NK₃ sites present in rat cerebral cortex membranes, 10 none of the compounds showed any significant affinities for these two sites (IC₅₀ $\geq 10^{5}$ nM).

In conclusion, this study has highlighted the importance of the basic secondary nitrogen, ortho substituent and aromatic moieties for NK_1 receptor binding affinity in this series of compounds. In addition, differences in the rank order of affinities for this series of compounds between rat and guinea-pig NK_1 binding sites confirm and extend recent studies which suggest the existence of species-dependent NK_1 receptor subtypes ${}^{1/4}, {}^{15}, {}^{16}$

Acknowledgement

The authors thank Dr Giles Ratcliffe, Mrs Lindsey Terry and Miss Elizabeth Allen for their technical assistance.

References and Notes

- Snider, R.M. et al.; Science 1991, 251, 435.
- 2. a) Nakanishi, S.; Physiol. Rev. 1987, 67, 117.
 - b) Maggio, J.E.; <u>Annu. Rev. Neurosci.</u> 1988, 11, 13.
 - c) Krause, J.E., MacDonald, M.R., Takeda, Y.; <u>BioEssays</u> 1989, 10, 62.
- 3. Von Euler, U.S., Gaddum, J.H.; <u>J. Physiol.</u> (London) 1932, 72, 74.

- 4. Chang, M.M., Leeman, S.E.; <u>J. Biol. Chem.</u> 1970, 245, 4784.
- 5. Guard, S., Watson, S.P.; Neurochem. Int. 1991, 18, 149.
- 6. Lowe, J.A. et al.; BioMed. Chem. Lett. 1991, 1, 129.
- 7. Lowe, J.A.; WO Patent 90/05525, 1990.
- 8. Warawa, E.J., Mueller, N.J., Jules, R.; <u>J. Med. Chem.</u> 1974, 17, 497.
- 9. Warawa, E.J., Mueller, N.J.; J. Med. Chem. 1975, 18, 587.
- 10 a) Lee, C.M., Campbell, N.J., William, B.J., Iversen, L.L.; <u>Eur.</u>
 <u>J. Pharmacol.</u> 1986, **130**, 209.
 - b) Guard, S., Watson, S.P., Maggio, J.E., Too, H.P., Watling, K.J.; Brit. J. Pharmacol. 1990, 99, 767.
- 11. Membranes (50-100 μ g protein/tube) were incubated (60 min at room temperature) with 10⁻¹⁰ M [125 I]-Bolton Hunter labelled substance P ([125 I]-BHSP) in a final volume of 300 μ l in the presence of 10^{-11} - 10^{-5} M test compound, and then rapidly filtered through Whatman GF/C filters using a Brandel Cell Harvester. Substance P (10^{-6} M) and the selective NK₁ receptor agonist [Sar 9 Met(O_2) 11] substance P (10^{-6} M), were used to define non-specific binding in rat and guinea-pig tissue, respectively. Data from 3-5 individual displacement curves were analyzed using an iterative curve fitting programme on RS1 and expressed as IC₅₀ values in nM (geometric mean -, + s.e.m.).
- 12. In <u>Substituent Constants for Correlation Analysis in Chemistry and Biology</u>, Hansch, C.; Leo, A.; Wiley-Interscience, New York, 1979.
- 13. Buck, S.H.; Shatzer, S.A.; Life Sci., 1988, 42, 2701.
- 14. Gitter, B.D. et al; Eur. J. Pharmacol 1991, 197, 237.
- 15. Watling, K.J. et al; Brit. J. Pharmacol. 1991, 104, 27P.
- 16. Beresford, I.J.M. et al; Brit. J. Pharmacol. 1991, 104, 292.